

Original Article

Antibiotic Resistance in *Enterococcus faecalis* Isolated from Hospitalized Patients

Esrafil Balaei Gajan^{1,2} • Adileh Shirmohammadi³ • Mohammad Aghazadeh⁴ • Mohammad Alizadeh⁵ • Alireza Sighari Deljavan^{6*} • Farzin Ahmadpour⁷

¹Dental and Periodontal Research Center, Tabriz University of Medical Sciences, Tabriz, Iran

²General Dentist, Department of Community Dentistry, Faculty of Dentistry, Tabriz University of Medical Sciences, Tabriz, Iran

³Associate Professor, Department of Periodontics, Faculty of Dentistry, Tabriz University of Medical Sciences, Tabriz, Iran

⁴Assistant Professor, Department of Microbiology, Faculty of Medicine, Tabriz University of Medical Sciences, Tabriz, Iran

⁵Lecture, Faculty of Nursery and Midwifery, Tabriz University of Medical Sciences, Tabriz, Iran

⁶Research Assistant, Faculty of Dentistry, Tabriz University of Medical Sciences, Tabriz, Iran

⁷Dental Student, Faculty of Dentistry, Tabriz University of Medical Sciences, Tabriz, Iran

*Corresponding Author; E-mail: alireza_sigharydeljavan@yahoo.com

Received: 2 November 2012; Accepted: 16 March 2013

J Dent Res Dent Clin Dent Prospect 2013;7(2):102-104 | doi: 10.5681/joddd.2013.018

This article is available from: <http://dentistry.tbzmed.ac.ir/joddd>

© 2013 The Authors; Tabriz University of Medical Sciences

This is an Open Access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/3.0>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Abstract

Background and aims. Enterococci are Gram-positive cocci that often occur in pairs (diplococci) or short chains. Beside developing high level of antibiotic resistance, these bacteria can cause wide range of disease in human, thus to help provide an effective treatment for infections caused by this genus, this study was conceived to provide information on *Enterococcus faecalis* Antibiotic resistance to widely used antibiotics in hospitalized patients.

Materials and methods. Disk diffusion agar and Broth dilution methods were used to perform Antibiogram test on isolated *Enterococcus faecalis*. Culture medium used for Disk diffusion agar test was Muller Hinton agar, and for Broth dilution methods, Muller Hinton broth culture medium was utilized. In disk diffusion agar method, different commercial antibiotics disks produced by Pharmaceutical companies were used. Microsoft Excel software was used to perform statistical analysis.

Results. Based on antibiograms of 105 cases, a high resistance to Synercid, Nalidixic acid, Oxacillin and Teofilin was detected whereas the lowest resistance observed in Nitrofurantoin, Vancomycin, Linezolid and Teicoplanin antibiotics.

Conclusion. According to the results, Teicoplanin, Vancomycin, Linezolid and Nitrofurantoin are recommended against *E. faecalis* species.

Key words: Antibiogram, *Enterococcus faecalis*, MIC.

Introduction

Enterococci are Gram-positive cocci that commensal inhabitants of the human intestine. Some strains of this genus have developed resistance to

antibiotics.¹ These bacteria are non-motile, without capsule and cultivated on the bile esculin agar (BEA) as a selective differential medium. *Enterococcus* grows fast in 37-42°C temperature and forms non-hemolytic colonies.² Members of the genus *Entero-*

coccus are capable of growing in the presence of 6.5% NaCl and 4% bile (oxgall) and hydrolyzing esculin to glucose and esculetin. Esculetin combines with ferric ions to produce a black complex.² Important clinical infections caused by *Enterococcus* include urinary tract infections, bacteremia, bacterial endocarditis, diverticulitis, and meningitis. They can tolerate 60°C heat for 30 minutes.³⁻⁵

This study was conceived to provide information on *Enterococcus faecalis* Antibiotic resistance to widely used antibiotics in hospitalized patients, considering dose and their type.

Materials and Methods

One-hundred and five samples were collected from patients hospitalized with enterococcus infections in Tabriz, Iran, from March 2012 to the end of June 2012.

The most simple and best methods for antibiogram are preparation tubular tenuities and disk diffusion agar. Bacterial suspensions with McFarland Standard 0.5% were cultured in Mueller-Hinton agar medium and impregnated paper disks were placed on the culture medium.^{2,3} 0.5% McFarland standard was prepared by mixing 0.05 ml of 1.175% barium chloride dihydrate ($\text{BaCl}_2 \cdot 2\text{H}_2\text{O}$), with 9.95 ml of 1% sulfuric acid (H_2SO_4). The mixture was confirmed by spectrophotometer at 625 nm wave length. The absorbency for 0.5% McFarland standard must be in 0.1-0.68 range. The standards are equivalent to $1-1.5 \times 10^8$ bacteria in milliliter.

The antibiotic disks used for antibiogram test included Vancomycin (30 mg), Nalidixic Acid (30 mg), Ampicillin (10 mg), Nitrofurantoin (300 mg), Oxacillin (1mg), Tetracycline (30 mg), Gentamicin (100 mg), Co-trimoxazole (1.25-22.75), Erythromycin (30 mg), Teicoplanin (30mg), Linezolid (30 mg), Ciprofloxacin, Synercid, Doxycycline.

After 24 hours of incubation in 37°C, the inhibition zone was measured with metric ruler and then matched with NCCLS (National Committee for Clinical Laboratory Standards) tables to finalize antimicrobial susceptibility tests.

Minimum inhibitory concentration (MIC) was used to determine antimicrobial resistance.

Antibiotic with different concentrations were prepared from antibiotic solution.^{1,6,7} 12 sterile tubes in the form of tower row triplet and added 1, 3 and 7 ml Muller Hinton broth culture environment to first, second, and third rows, respectively. Then, we added 1 ml of antibiotic solution to three first rows. 3 ml was taken from the third tube and added 1 ml to second row of triplet tube. After mixing, 3 ml was taken from sixth tube again, and 1 ml was added to the third row of triplet tubes. Again 3 ml was taken from ninth tube and 1 ml was added to the fourth row of triplet tubes. Accordingly, antibiotic concentrations were prepared in tubes 1 to 12 as 1024, 512, 256, 128, 64, 32, 16, 8, 4, 2, 1, 0.5, and 0.25, respectively. 1 ml of each concentrations was poured in the 12 sterile tubes and then bacterium suspensions were added (0.5 McFarland 100 land and 19.9 ml Muller Hinton broth environment). There are 1.5×10^8 bacteria in each milliliter, and 1 ml was added to each of the 12 tubes. Then, the tubes were placed in incubator at 37°C for 24 hours.^{6,8} The development of opacity in the tubes indicated bacterial resistance. MIC is the case antibiotic.^{8,9} To determine resistance percentage and distinguish multi-resistance lineage of *Enterococcus faecalis* isolated from patients Micro-soft Excel software was used.

Results

Based on antibiograms of 105 subjects, the presence of antibiotic-resistance was more prevalently observed with Tetracycline, Oxacillin and Nalidixic Acid. Also, a 100% resistance was observed in the case of Synercid. Percentage of enterococcus antibiotic-resistance to different antibiotics is shown in the Table 1.

Discussion

For effective treatment of bacterial infections, adequate concentration of antibiotic must be present in the site of infection. This concentration must show either bactericidal or bacteriostatic activity against susceptible bacteria. Therefore, if in a case, the level of antibiotic required for preventing bacteria from growing, exceeds toxic concentrations, the bacterium

Table 1. Percentage of resistant enterococci to different antibiotics

Antibiotics	Synercid	Nalidixic acid	Oxacillin	Teicoplanin	Ciprofloxacin	Doxycycline	Ampicillin	Gentamycin	Erythromycin	Amikacin	chloramphenicol	Nitrofurantoin	Linezolid	Vancomycin	Teicoplanin
Resistant (%)	100	94	93	83	44	44	4	36.2	29	27	2	18.6	5.1	3.6	3.3

is considered antibiotic-resistant.

Enterococcus faecalis is Gram-positive cocci that often in the form of diplococci or short chains. *E. faecalis* can cause endocarditis and bacteremia, urinary tract infections (UTI), meningitis, and other infections in humans. This species can grow in bile esculin agar and environments with 6.5% NaCl. It grows fast at 37-42°C temperature while it can tolerate 60°C heat for 30 minutes.

Recent reports show that resistance of these bacteria to commonly used antibiotics is increasing worldwide. Even Vancomycin-resistant and Gentamicin-resistant species of *E. faecalis* are reported.^{10,11} An antibiogram test is, hence, required for effective treatment. In the present study, the highest rate of resistance was seen with Synercid (100%). For Nalidixic acid, resistant was 94% and for Oxacillin it was about 93%. Also about 83% of cases showed resistance to Teofilin which is regarded a high resistance. These antibiotics cannot be appropriate prescriptions for the treatment of *E. faecalis* infections. Moderate resistance was reported in Ciprofloxacin, Doxycycline, Amikacin, Gentamycin, and the least percentage of resistance was obtained with Teicoplanin, Vancomycin, Linezolid, and Nitrofurantoin.

According to the results, it can be concluded that instead of prescribing antibiotics like Gentamycin, Amikacin or Oxacillin against *E. faecalis* infection, antibiotics like Teicoplanin, Vancomycin, Linezolid, and Nitrofurantoin are recommended along with antibiotics like chloramphenicol and Ampicillin as the second-line agents.

References

1. Jawetz E, Melnick JLL, Adelberg EA. *Medical Microbiology LANGE*, 21st ed. California: Mc Grow Hill; 2001. p. 43-8.
2. Forbes BA, Sahm DF, Weissfeld AS. *Bailey & Scott's Diagnostic Microbiology*, 12th ed. St Louis: Mosby; 2007. p. 101-10.
3. Shankar N, Lockett CV, Baghdayan AS, Drachenberg C, Gilmore MS, Johnson DE. Role of *Enterococcus faecalis* surface protein Esp in the pathogenesis of ascending urinary tract infection. *Infect Immun* 2001;69:4366-72.
4. Benca J, Ondrusova A, Huttova M, Rudinsky B, Kisac P, Bauer F. Neuroinfections due to *Enterococcus faecalis* in children. *Neuro Endocrinol Lett* 2007;28:32-3.
5. Zoletti GO, Pereira EM, Schuenck RP, Teixeira LM, Siqueira JF Jr, dos Santos KR. Characterization of virulence factors and clonal diversity of *Enterococcus faecalis* isolates from treated dental root canals. *Res Microbiol* 2011;162:151-8.
6. Grant SG, Jessee J, Bloom FR, Hanahan D. Differential plasmid rescue from transgenic mouse DNAs into *Escherichia coli* methylation-restriction mutants. *Proc Natl Acad Sci USA* 1990;87:4645-9.
7. Marrag PR, Rosental KS, Kobayashi GS, Pfaller MA. *Medical Microbiology*, 4th ed. St Louis: Mosby; 2002. p. 45-8.
8. Courvalin P. Vancomycin resistance in gram-positive cocci. *Clin Infect Dis* 2006;42:525-34.
9. Kousedghi H, Ahangari Z, Eslami G, Ayatollahi A. Antibacterial activity of propolis and Ca (OH) 2 against *Lactobacillus*, *Enterococcus faecalis*, *Peptostreptococcus* and *Candida albicans*. *African Journal of Microbiology Research* 2012;14:3510-5.
10. Aligholi M, Emaneini M, Taherikalani M, Shahsavan S, Jabalameli F, Asadollahi P, et al. Time-kill study and synergistic activity of cell-wall inhibitor antibiotics in combination with gentamicin against *Enterococcus faecalis* and *Enterococcus faecium*. *Acta Microbiol Immunol Hung* 2011;58:219-26.
11. Furustrand Taffin U, Majic I, Zalila Belkhodja C, Betrisey B, Corvec S, Zimmerli W, et al. Gentamicin improves the activities of daptomycin and vancomycin against *Enterococcus faecalis* in vitro and in an experimental foreign-body infection model. *Antimicrob Agents Chemother* 2011;55:4821-7.